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**SUPRAMEDULLARY MODULATION OF SYMPATHETIC
VASOMOTOR FUNCTION**

Anthony JM Verberne, William Lam, Neil C Owens and Daniela Sartor

*University of Melbourne, Department of Medicine, Clinical Pharmacology and Therapeutics Unit, Austin
& Repatriation Medical Centre, Heidelberg, Victoria, Australia*

SUMMARY

1. **Supramedullary structures including the ventral medial prefrontal cortex (MPFC) and the midbrain cuneiform nucleus (CnF) project directly and indirectly to premotor sympathoexcitatory neurons of the rostral ventrolateral medulla (RVLM) that are critically involved in the generation of sympathetic vasomotor tone.**

2. **Electrophysiological studies have demonstrated that activation of depressor sites within the MPFC is associated with splanchnic sympathetic vasomotor inhibition and inhibition of the activity of RVLM sympathoexcitatory neurons.**

3. **Antidromic mapping and anatomical studies support the notion that a relay in the nucleus tractus solitarius is involved in the cardiovascular response to MPFC stimulation.**

4. **The midbrain CnF, which lies adjacent to the midbrain periaqueductal grey, is a sympathoexcitatory region of the midbrain reticular formation. Sympathoexcitatory responses evoked from the CnF are associated with short-latency excitation of RVLM neurons.**

5. **Cuneiform nucleus stimulation induces the expression of mRNA for the immediate early genes *c-fos* and *NGFI-A* in midbrain, pontine and hypothalamic structures.**

6. **The MPFC and CnF are supramedullary structures with opposing modulatory influences on sympathetic vasomotor drive, whose roles in cardiovascular control mechanisms warrant further investigation.**

Key words: blood pressure, cardiovascular system, rostral ventrolateral medulla, sympathetic vasomotor outflow.

INTRODUCTION

In recent years the focus of central cardiovascular control research has been on the origins of sympathetic vasomotor tone and the

neurocircuitry associated with reflex circulatory control.^{1–4} The importance of spinally projecting neurons within the rostral ventrolateral medulla (RVLM) in the generation of sympathetic vasomotor tone has been established in a number of species,^{3,5–7} so that these neurons are generally regarded as premotor sympathoexcitatory neurons that provide tonic drive to the sympathetic preganglionic vasomotor neurons in the thoracolumbar regions of the spinal cord. In addition, an increasing body of evidence suggests that the central nervous system is capable of generating highly differentiated patterns of sympathetic vasomotor activity resulting in regionally specific changes in blood flow,^{8,9} rather than a global activation of the sympathetic nervous system, as first postulated by Cannon.¹⁰ Differentiated patterns of regional sympathetic vasomotor outflow are probably expressed through activation of specific subgroups of RVLM sympathoexcitatory neurons committed to specific groups of sympathetic preganglionic neurons which innervate appropriate peripheral vascular beds. It is unlikely that the RVLM itself integrates the specific sensory inputs that lead to selected patterns of regional sympathetic response; this is likely to be the role of higher brain structures that have access to sensory information relevant to the behavioural state of the organism. For example, it has been repeatedly demonstrated that electrical and chemical stimulation of regions within the midbrain periaqueductal grey area (PAG) or the hypothalamus leads to a differential sympathetic vasomotor response and pattern of regional vascular response.^{11–17} Presumably, neurons in the PAG and hypothalamus are capable of activating specific subgroups of RVLM sympathoexcitatory neurons by virtue of direct connections with the RVLM.^{16,18–20}

**SUPRAMEDULLARY STRUCTURES THAT
INFLUENCE SYMPATHETIC VASOMOTOR
TONE**

There are a number of supramedullary regions other than the PAG and hypothalamus that are capable of influencing circulatory function (for a recent review see Dampney²). In the present review we will focus on two supramedullary structures that are known to have opposing influences on arterial blood pressure (BP) and sympathetic vasomotor activity: the depressor area of the medial prefrontal cortex (MPFC) and the cuneiform nucleus (CnF), a sympathoexcitatory area of the lateral midbrain reticular formation (Fig. 1).

Correspondence: Dr AJM Verberne, University of Melbourne, Department of Medicine, Clinical Pharmacology and Therapeutics Unit, Austin & Repatriation Medical Centre, Heidelberg, Victoria 3084, Australia. Email: <tonyv@austin.unimelb.edu.au>

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MEDIAL PREFRONTAL CORTEX: A CORTICAL SYMPATHOINHIBITORY REGION

The medial prefrontal cortex (MPFC; Fig. 1) has been defined as the cortical region that receives afferent input from the mediodorsal thalamic nucleus (MD)^{21,22} and is subdivided into the prelimbic (PL) and infralimbic (IL) areas. The PL area receives afferents predominantly from the medial and lateral MD,²³ while the IL area does not appear to be a target of these projections.²² The MPFC and MD are connected reciprocally^{21,24–26} and the input to the MPFC from the MD is probably glutamatergic.²⁷

The MPFC has been termed a 'visceromotor' cortical area.^{28,29} Whether the MPFC has any special claim to this terminology is uncertain as a number of other, albeit linked, cortical regions also influence visceral motor function. Nevertheless, the MPFC contains neurons that project to a number of other brain regions associated with visceral motor control or visceral afferent processing and its activation leads to various changes in the activity of effector organs and tissues influenced by the autonomic nervous system.²⁸ Thus, electrical stimulation of this region elicits changes in arterial BP,^{30–32} heart rate, respiration, gastric motility^{28,33} and potentially micturition, as the MPFC projects to Barrington's nucleus, the pontine micturition centre.³⁴ Electrical stimulation of the MPFC usually elicits depressor responses in the anaesthetized rat, although pressor responses have been reported when the MPFC is stimulated in awake animals.³⁰ Cechetto and Saper³⁵ reviewed the literature on the role of the MPFC in cardiovascular control and since then we and other researchers have sought additional evidence for such a role and for the mechanisms associated with MPFC stimulation-evoked depressor responses.

PATHWAYS OF THE CORTICAL SYMPATHOINHIBITORY SYSTEM

The pathway(s) that mediate the depressor response elicited by MPFC stimulation have not been completely elucidated. A potential candidate pathway was identified when a number of tract-tracing studies identified direct projections from the MPFC to brainstem structures associated with autonomic function.^{36–38} Neuroanatomical studies performed in rats, rabbits and cats have demonstrated that the nucleus tractus solitarius (NTS) in the dorsomedial medulla receives afferents not only from peripheral visceral sources, but also from supramedullary structures, including the MPFC and the insular cortex (IC)^{29,36–45} (Fig. 1). These observations led to speculation that the cardiovascular (and/or other autonomic) responses evoked by microstimulation of the MPFC may be mediated by a projection to the NTS. As yet, there is no unequivocal evidence that cortical neurons projecting to the NTS actually synapse with NTS neurons involved in cardiovascular control. Nevertheless, there is considerable overlap between the sites within the NTS that receive both cortical and cardiopulmonary visceral afferents,^{36,46} suggesting that cortical neurons projecting to the NTS may modulate visceral reflex function.

Extracellular single-unit recordings made from the depressor area of the MPFC have demonstrated that neurons in this area may be activated antidromically by stimulation within the depressor area of the NTS.^{47,48} Using antidromic mapping techniques,⁴⁹ our laboratory has demonstrated that axons of MPFC neurons terminate within a depressor area of the caudal/intermediate NTS.⁴⁸ These observations raise the possibility that cortical inputs to the NTS may be involved

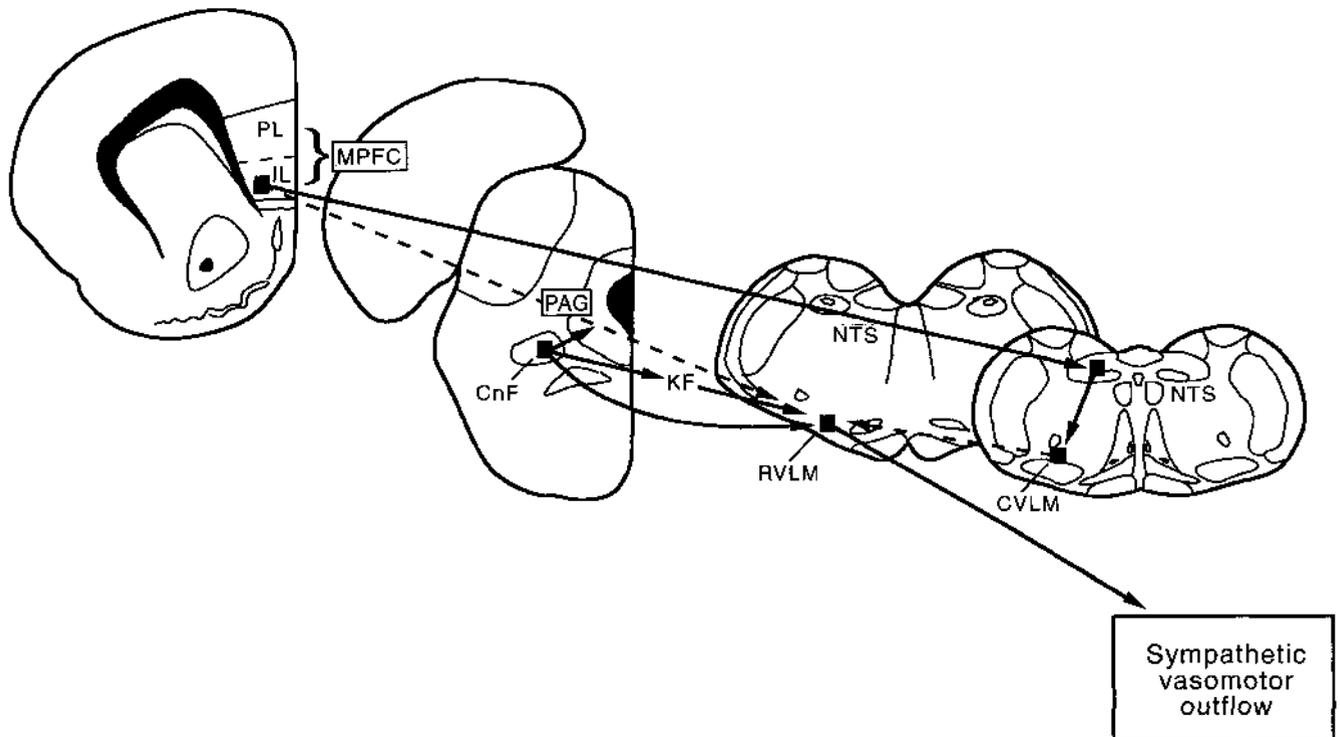


Fig. 1. Anatomical location and projections of the medial prefrontal cortex (MPFC) and midbrain cuneiform nucleus (CnF) to pontomedullary structures associated with central cardiovascular control mechanisms. CVLM, caudal ventrolateral medulla; IL, infralimbic area; KF, Kölliker-Fuse nucleus; NTS, nucleus tractus solitarius; PAG, periaqueductal grey area; PL, prelimbic area; RVLM, rostral ventrolateral medulla. Solid lines represent excitatory pathways; dashed lines represent inhibitory pathways.

in modulation of the baroreceptor reflex. Indeed, it has been demonstrated that excitotoxic lesions of the MPFC reduce the gain of the heart rate baroreflex in normotensive but not in spontaneously hypertensive rats.^{28,50,51}

Electrical stimulation of the IL depressor area results in the appearance of Fos-like immunoreactivity in neurons in a number of brain regions known to receive IL afferents, including the NTS and the caudal ventrolateral medulla at the level of the obex (D Owens *et al.*, unpubl. obs., 1996). This suggests that the projection to the NTS is excitatory and is consistent with activation of baroreceptor-related NTS neurons and the production of a depressor response through activation of the baroreflex arc (Fig. 2).⁵² It is interesting to note that IC afferents to the NTS are probably also excitatory.⁵³

Alternatively, a recent preliminary report indicates that at least part of the MPFC stimulation-evoked depressor response is associated with a relay in the lateral hypothalamic area.⁵⁴ Similarly, it has been suggested that neurons within the lateral hypothalamic area participate in the depressor response elicited from the lateral prefrontal cortex, the rostral extension of the IC.^{55,56} It is therefore conceivable that the MPFC stimulation-evoked depressor response is mediated by a number of separate pathways that include both hypothalamic as well as medullary relays (Fig. 2).

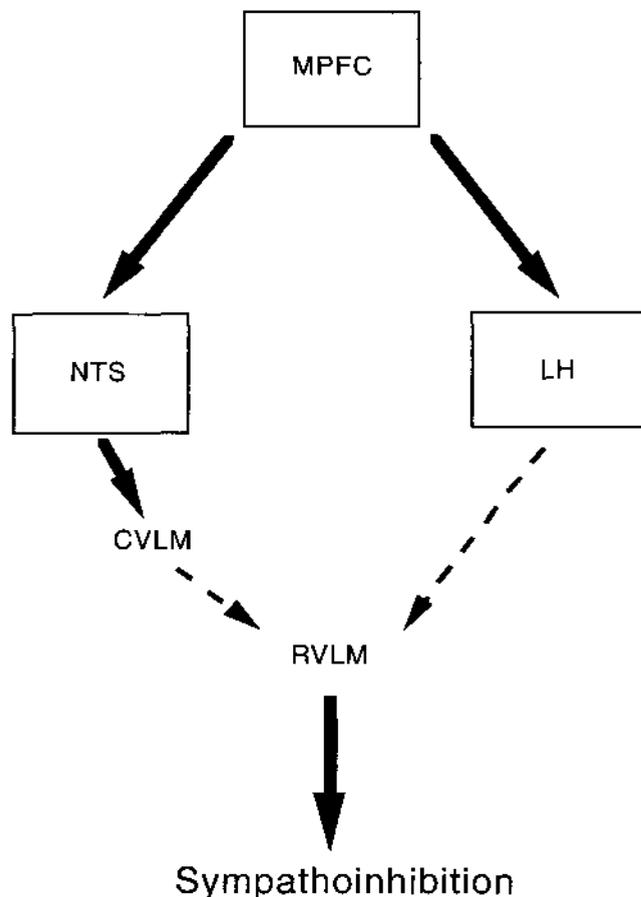


Fig. 2. Potential pathways mediating medial prefrontal cortex (MPFC) stimulation-evoked depressor and sympathoinhibitory responses. CVLM, caudal ventrolateral medulla; LH, lateral hypothalamic area; NTS, nucleus tractus solitarius; RVLM, rostral ventrolateral medulla. Solid lines represent excitatory pathways; dashed lines represent inhibitory pathways.

In a recent study, our laboratory examined the influence of stimulation of the MPFC depressor region on BP, sympathetic vasomotor discharge and the discharge of the premotor sympathoexcitatory neurons of the RVLM.⁵⁷ It was demonstrated that depressor responses elicited from the MPFC are associated with inhibition of the sympathetic vasomotor outflow, particularly to the abdominal visceral vasculature innervated by the splanchnic nerve.⁵⁷ These depressor and sympathoinhibitory responses were elicited from a wide range of dorsoventral locations within the MPFC. This is possibly due to the fact that MPFC neurons projecting to structures such as the NTS are located in both the PL and IL areas, although they are found predominantly in the latter.²⁸ Electrical stimulation of the MPFC also resulted in the inhibition of some RVLM sympathoexcitatory neurons.⁵⁷ These neurons are profoundly barosensitive and are considered to be critical in the generation of sympathetic vasomotor tone.^{3,58} This suggests that MPFC stimulation-evoked depressor and sympathoinhibitory responses are mediated, at least in part, by inhibition of RVLM premotor, sympathoexcitatory neurons. Although a direct projection from the MPFC to the area of the RVLM containing premotor sympathoexcitatory neurons has been reported previously,^{42,59} this projection appears to be relatively sparse and, therefore, may not have a major role in mediating cortically evoked depressor responses. Alternatively, activation of the MPFC probably results in inhibition of the activity of RVLM neurons via a polysynaptic route that may include relays in the NTS and/or the hypothalamus.

In summary, the MPFC contains a depressor and sympathoinhibitory region whose role in central cardiovascular control mechanisms remains obscure. However, it is probable that the MPFC is involved in the mechanisms that lead to coordination of circulatory function with ongoing behaviour.

CUNEIFORM NUCLEUS: A MIDBRAIN PRESSOR AND SYMPATHOEXCITATORY REGION

The cuneiform nucleus is situated lateral to the midbrain trigeminal nucleus adjacent to the caudal lateral PAG area.⁶⁰ Anatomical investigations have demonstrated a projection from the CnF to a number of structures that are associated with cardiorespiratory mechanisms.⁶⁰ Furthermore, a projection from CnF to the raphe magnus nucleus, an area involved in a descending pain inhibitory system, has also been described previously.⁶⁰⁻⁶² The CnF projects to the PAG and some of these neurons may be glutamatergic.^{63,64} These connections suggest that the CnF may be involved in some way with the central circuits associated with endogenous pain control systems.⁶⁵

The PAG has also been implicated in central circulatory control,⁶⁰ where it is considered to be critical for the expression of the autonomic and behavioural components of the defence response.^{16,20} Similarly, it has been suggested that the CnF could be involved in a form of defensive behavioural and cardiovascular response to threatening or painful stimuli.⁶⁶ Chemical and electrical stimulation of the superior colliculus also results in cardiovascular responses similar to CnF stimulation⁶⁷ and it has been speculated that, as the superior colliculus receives largely visual input, a pathway involving the superior colliculus, the CnF and other, as yet unidentified structures, may be involved in mediating defence responses associated with threatening visual stimuli.⁶⁷

PATHWAYS OF THE CnF SYMPATHOEXCITATORY SYSTEM

It is conceivable that activation of the CnF may excite neurons in the PAG which, in turn, project to the premotor, sympathoexcitatory neurons of the RVLM.^{16,20,59} A direct projection from the CnF to the sympathoexcitatory area of the RVLM has been reported⁶⁰ but appears to be relatively sparse. While this does not necessarily preclude its involvement in the cardiovascular responses elicited from the CnF, earlier retrograde tracing studies from the RVLM have not reported a similar projection.⁵⁹

It has also been postulated that the cardiovascular responses elicited from the CnF may be mediated by its much stronger connections with the Kölliker-Fuse nucleus, which projects strongly to the RVLM.⁶⁰ Electrical stimulation of both of these areas leads to elevation of arterial BP.^{60,68,69} Our laboratory has examined the mechanism of the sympathoexcitatory response elicited by electrical stimulation of the CnF using: (i) electrophysiological techniques, such as extracellular single unit recording; and (ii) molecular biological techniques, such as *in situ* hybridization histochemical detection of immediate early gene (IEG) mRNA as an index of neuronal activation.⁷⁰ Electrical stimulation of the CnF elicited a pressor response accompanied by excitation of the lumbar sympathetic vasomotor discharge and activation of RVLM sympathoexcitatory vasomotor neurons.⁷¹ The peak latency for excitation of RVLM sympathoexcitatory neurons by CnF stimulation (16 ± 1 ms) approximates the difference between the earliest peak latency of the lumbar sympathoexcitatory response elicited by CnF stimulation

(102 ± 3 ms) and the peak latency of the lumbar sympathoexcitatory response elicited by RVLM stimulation (81 ± 1 ms; Fig. 3b).¹⁶ These observations are consistent with the hypothesis that the RVLM relays CnF stimulation-evoked sympathoexcitatory responses.

Rostral ventrolateral medulla premotor sympathoexcitatory neurons have been subdivided into two major groups based on conduction velocity of the spinally projecting axon and sensitivity to catecholamines.^{3,72} Electrical stimulation of the RVLM elicits a bimodal sympathoexcitatory response (two peaks of sympathoexcitation with different latencies), probably as a result of activation of the medullospinal neurons with different conduction velocities.⁷³ Interestingly, stimulation of the CnF produced a bimodal sympathoexcitatory response with latencies that are compatible with activation of the RVLM premotor sympathoexcitatory neurons with different axonal spinal conduction velocities.^{71,74} An additional or alternative explanation for the late peak of sympathoexcitation may also be that CnF stimulation excites slow-conducting cells within the raphe pallidus nucleus, which may have a sympathoexcitatory function.⁷⁵

Neuroanatomical studies indicate that the CnF projects only weakly to the RVLM.^{59,60} This suggests that the pathway from the CnF mediating sympathoexcitation may be polysynaptic and involves a relay located elsewhere (Fig. 3). A potential candidate may be the Kölliker-Fuse nucleus of the dorsolateral pons, which receives afferents from the CnF.⁶⁰ Studies that have examined the IEG response to CnF stimulation have supported this notion.^{76,77} Cuneiform nucleus stimulation produced significant increases in the levels of

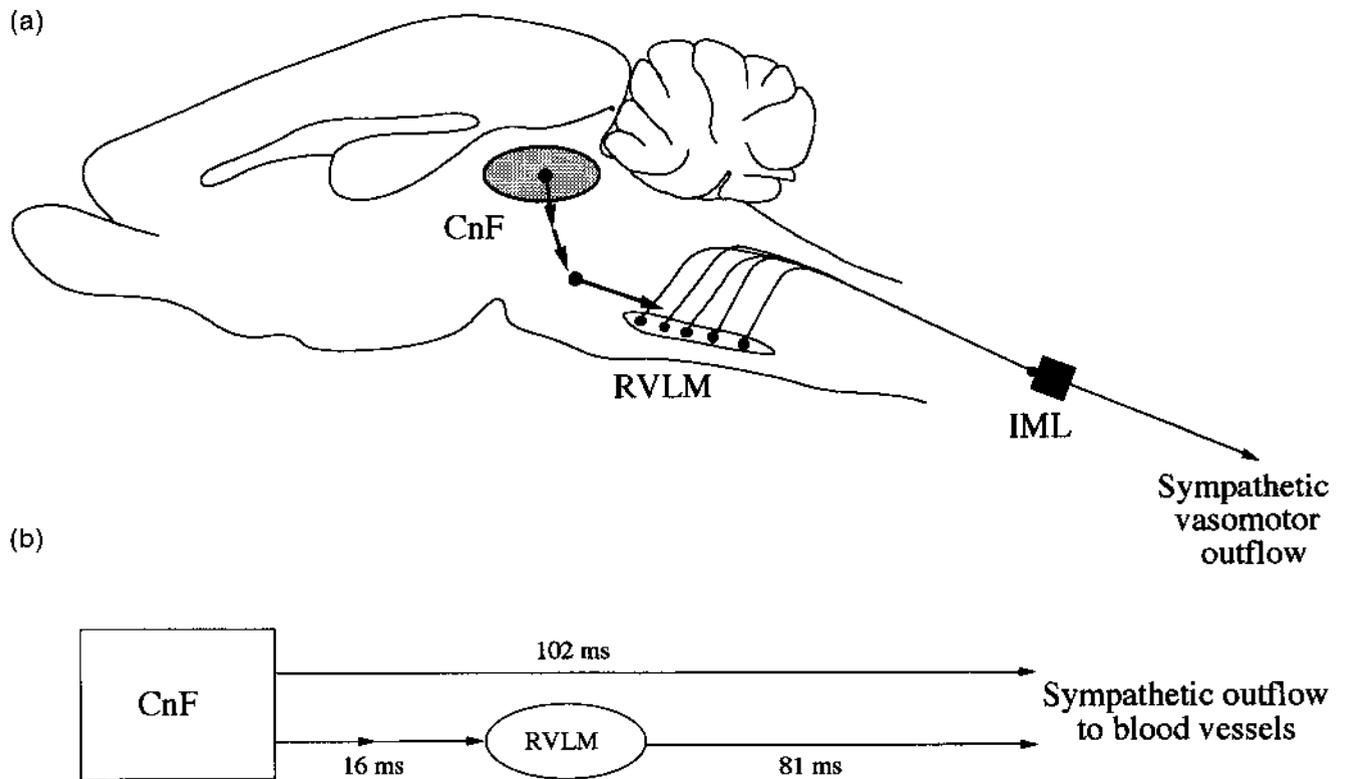


Fig. 3. (a) Cuneiform nucleus (CnF) sympathoexcitatory pathway. Activation of the CnF produces activation of the sympathetic vasomotor outflow by activation of premotor, sympathoexcitatory neurons of the rostral ventrolateral medulla (RVLM). This is most probably a polysynaptic input to the RVLM. (b) Electrical stimulation of the CnF activates RVLM sympathoexcitatory neurons (latency = 16 ms) and sympathetic vasomotor outflow (latency = 102 ms). See text for discussion.

IEG mRNA (*c-fos* and *NGFI-A*, nerve growth factor inducible form, type A) in the Kölliker-Fuse nucleus and parabrachial complex ipsilaterally and the dorsal PAG area bilaterally. In addition, various forebrain structures, such as the ventromedial, dorsomedial and lateroanterior hypothalamic nuclei and lateral and anterior hypothalamic areas, displayed significantly elevated levels of IEG mRNA.⁷⁶

At present, the role of the CnF in central cardiovascular control is uncertain. The limited evidence available suggests that the CnF may be involved in the autonomic and behavioural components of defensive responses to painful or threatening stimuli.⁶⁶

In summary, stimulation of the CnF produces elevations of arterial BP and sympathetic vasomotor activity, at least in part, by activating RVLM sympathoexcitatory vasomotor neurons. Furthermore, the CnF appears to be an additional supramedullary site from which sympathetic vasomotor responses may be elicited but whose precise role in central cardiovascular control remains to be elucidated.

POTENTIAL FUNCTIONAL ROLES OF THE MPFC AND CnF IN CENTRAL CARDIOVASCULAR CONTROL MECHANISMS

Supramedullary structures with anatomical connections to structures known to be involved in central cardiovascular control mechanisms are firmly established and are probably important for the generation of patterns of circulatory response and adjustment in concert with behavioural change. It has been suggested previously that the MPFC may be involved in the termination of defensive behaviour⁷⁸ or in the development of habituation to stimuli, which induce defence-like responses. The involvement of the CnF in nociceptive mechanisms⁷⁹ suggests that it may be implicated in the cardiovascular responses to painful or threatening stimuli.⁶⁶ In addition, the CnF appears to coincide with the so-called midbrain locomotor region, which is active during exercise⁸⁰ and so may participate in the cardiovascular response observed during physical activity.

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REFERENCES

- Chalmers JP, Kapoor V, Llewellyn-Smith IJ, Minson JB, Pilowsky PM. Central control of blood pressure. *Eur. Heart J.* 1992; **13**: 2–9.
- Dampney RAL. Functional organization of central pathways regulating the cardiovascular system. *Physiol. Rev.* 1994; **74**: 321–63.
- Guyenet PG. Role of the ventral medulla oblongata in blood pressure regulation. In: Loewy AD, Spyer KM (eds). *Central regulation of autonomic functions*. Oxford University Press, New York. 1990; 145–67.
- Kumada M, Terui N, Kuwaki T. Arterial baroreceptor reflex: Its central and peripheral neural mechanisms. *Prog. Neurobiol.* 1990; **35**: 331–61.
- McAllen RM, May CN, Shafiq AD. Functional anatomy of sympathetic premotor cell groups in the medulla. *Clin. Exp. Hypertens.* 1995; **17A**: 209–21.
- Dormer KJ, Anwar M, Ashlock SR, Ruggiero DA. Organization of presumptive catecholamine-synthesizing neurons in the canine medulla oblongata. *Brain Res.* 1993; **601**: 41–64.
- Terui N, Saeki Y, Kumada M. Barosensory neurons in the rostral ventrolateral medulla mediate the renal sympathetic reflex in rabbits. *Clin. Exp. Hypertens.* 1988; **1A**: 269–74.
- Dampney RA, Goodchild AK, McAllen RM. Vasomotor control by subretrofacial neurones in the rostral ventrolateral medulla. *Can. J. Physiol. Pharmacol.* 1987; **65**: 1572–9.
- Dampney RA, McAllen RM. Differential control of sympathetic fibres supplying hindlimb skin and muscle by subretrofacial neurones in the cat. *J. Physiol.* 1988; **395**: 41–56.
- Cannon WB. *Bodily Changes in Pain, Hunger, Fear and Rage*, 2nd edn. Appleton, New York. 1929.
- Lovick TA. Ventrolateral medullary lesions block the antinociceptive and cardiovascular responses elicited by stimulating the dorsal periaqueductal grey matter in rats. *Pain* 1985; **21**: 241–52.
- Carrive P. The periaqueductal gray and defensive behavior: Functional representation and neuronal organization. *Behav. Brain Res.* 1993; **58**: 27–47.
- Carrive P, Bandler R. Viscerotopic organization of neurons subserving hypotensive reactions within the midbrain periaqueductal grey: A correlative functional and anatomical study. *Brain Res.* 1991; **541**: 206–15.
- Carrive P, Bandler R, Dampney RA. Viscerotopic control of regional vascular beds by discrete groups of neurons within the midbrain periaqueductal grey. *Brain Res.* 1989; **493**: 385–90.
- Verberne AJ, Struyker Boudier HA. Midbrain central grey: Regional haemodynamic control and excitatory amino acidergic mechanisms. *Brain Res.* 1991; **550**: 86–94.
- Verberne AJ, Guyenet PG. Midbrain central gray: Influence on medullary sympathoexcitatory neurons and the baroreflex in rats. *Am. J. Physiol.* 1992; **263**: R24–33.
- Yardley CP, Hilton SM. The hypothalamic and brainstem areas from which the cardiovascular and behavioural components of the defence reaction are elicited in the rat. *J. Auton. Nerv. Syst.* 1986; **15**: 227–44.
- Li P, Lovick TA. Excitatory projections from hypothalamic and midbrain defense regions to nucleus paragigantocellularis lateralis in the rat. *Exp. Neurol.* 1985; **89**: 543–53.
- Luiten PG, ter Horst GJ, Steffens AB. The hypothalamus, intrinsic connections and outflow pathways to the endocrine system in relation to the control of feeding and metabolism. *Prog. Neurobiol.* 1987; **28**: 1–54.
- Carrive P, Bandler R, Dampney RA. Anatomical evidence that hypertension associated with the defence reaction in the cat is mediated by a direct projection from a restricted portion of the midbrain periaqueductal grey to the subretrofacial nucleus of the medulla. *Brain Res.* 1988; **460**: 339–45.
- Rose JE, Woolsey CN. Structure and relations of limbic cortex and anterior thalamic nucleus in rabbit and cat. *J. Comp. Neurol.* 1948; **89**: 279–347.
- Krettek JE, Price JL. The cortical projections of the mediodorsal nucleus and adjacent thalamic nuclei in the rat. *J. Comp. Neurol.* 1977; **171**: 157–92.
- Conde F, Audinat E, Maire-Lepoivre E, Crepel F. Afferent connections of the medial frontal cortex of the rat. A study using retrograde transport of fluorescent dyes. I. Thalamic afferents. *Brain Res. Bull.* 1990; **24**: 341–54.
- Leonard CM. The prefrontal cortex of the rat. I. Cortical projection of the mediodorsal nucleus. II. Efferent connections. *Brain Res.* 1969; **12**: 321–43.
- Beckstead RM. An autoradiographic examination of cortico-cortical and subcortical projections of the mediodorsal-projection (prefrontal) cortex of the rat. *J. Comp. Neurol.* 1979; **184**: 43–62.
- Martinez-Moreno E, Llamas A, Avendano C, Renes E, Reinoso-Suarez F. General plan of the thalamic projections to the prefrontal cortex in the cat. *Brain Res.* 1987; **407**: 17–26.
- Pirot S, Jay TM, Glowinski J, Thierry AM. Anatomical and electrophysiological evidence for an excitatory amino acid pathway from the thalamic mediodorsal nucleus to the prefrontal cortex in the rat. *Eur. J. Neurosci.* 1994; **6**: 1225–34.

28. Neafsey EJ. Prefrontal cortical control of the autonomic nervous system: Anatomical and physiological observations. In: Uylings HBM, van Eden CG, De Bruin JPC, Corner MA, Feenstra MGP (eds). *Progress in Brain Research*. Elsevier, Amsterdam. 1990; 147–65.
29. Neafsey EJ, Hurley-Gius KM, Arvanitis D. The topographical organization of neurons in the rat medial frontal, insular and olfactory cortex projecting to the solitary nucleus, olfactory bulb, periaqueductal gray and superior colliculus. *Brain Res.* 1986; **377**: 561–70.
30. Burns SM, Wyss JM. The involvement of the anterior cingulate cortex in blood pressure control. *Brain Res.* 1985; **340**: 71–7.
31. Hardy SGP, Holmes DE. Prefrontal stimulus-produced hypotension in rat. *Exp. Brain Res.* 1988; **73**: 249–55.
32. Powell DA, Watson K, Maxwell B. Involvement of subdivisions of the medial prefrontal cortex in learned cardiac adjustments in rabbits. *Behav. Neurosci.* 1994; **108**: 294–307.
33. Hurley-Gius KM, Neafsey EJ. The medial frontal cortex and gastric motility: Microstimulation results and their possible significance for the overall pattern of organization of rat frontal and parietal cortex. *Brain Res.* 1986; **365**: 241–8.
34. Valentino RJ, Page ME, Luppi PH, Zhu Y, Van Bockstaele E, Aston-Jones G. Evidence for widespread afferents to Barrington's nucleus, a brainstem region rich in corticotropin-releasing hormone neurons. *Neuroscience* 1994; **62**: 125–43.
35. Cechetto DF, Saper CB. Role of the cerebral cortex in autonomic function. In: Loewy AD, Spyer KM (eds). *Central regulation of autonomic functions*. Oxford University Press, New York. 1990; 208–23.
36. van der Kooy D, Koda LY, McGinty JF, Gerfen CR, Bloom FE. The organization of projections from the cortex, amygdala, and hypothalamus to the nucleus of the solitary tract in rat. *J. Comp. Neurol.* 1984; **224**: 1–24.
37. Terreberry RR, Neafsey EJ. Rat medial frontal cortex: A visceral motor region with a direct projection to the solitary nucleus. *Brain Res.* 1983; **278**: 245–9.
38. Terreberry RR, Neafsey EJ. The rat medial frontal cortex projects directly to autonomic regions of the brainstem. *Brain Res. Bull.* 1987; **19**: 639–49.
39. Buchanan SL, Thompson RH, Maxwell BL, Powell DA. Efferent connections of the medial prefrontal cortex in the rabbit. *Exp. Brain Res.* 1994; **100**: 469–83.
40. Willett CJ, Gwyn DG, Rutherford JG, Leslie RA. Cortical projections to the nucleus of the tractus solitarius: An HRP study in the cat. *Brain Res. Bull.* 1986; **16**: 497–505.
41. Yasui Y, Itoh K, Shigemoto R, Mizuno N. Topographical projections from the cerebral cortex to the nucleus of the solitary tract in the cat. *Exp. Brain Res.* 1991; **85**: 75–84.
42. Hurley KM, Herbert H, Moga MM, Saper CB. Efferent projections of the infralimbic cortex of the rat. *J. Comp. Neurol.* 1991; **308**: 49–276.
43. Ba M'hamed S, Sequeira H, Poulain P, Bennis M, Roy JC. Sensorimotor cortex projections to the ventrolateral and dorsomedial medulla oblongata in the rat. *Neurosci. Lett.* 1993; **164**: 195–8.
44. van der Kooy D, McGinty JF, Koda LY, Gerfen CR, Bloom FE. Visceral cortex: A direct connection from prefrontal cortex to the solitary nucleus in rat. *Neurosci. Lett.* 1982; **33**: 123–7.
45. Takagishi M, Chiba T. Efferent projections of the infralimbic (area 25) region of the medial prefrontal cortex in the rat: An anterograde tracer PHA-L study. *Brain Res.* 1991; **566**: 26–39.
46. Ruggiero DA, Mraovitch S, Granata AR, Anwar M, Reis DJ. A role of insular cortex in cardiovascular function. *J. Comp. Neurol.* 1987; **257**: 189–207.
47. Ruit KG, Neafsey EJ. Hippocampal input to a 'visceral motor' corticobulbar pathway: An anatomical and electrophysiological study in the rat. *Exp. Brain Res.* 1990; **82**: 606–16.
48. Owens NC, Verberne AJM. An electrophysiological study of the medial prefrontal cortical projection to the nucleus of the solitary tract in rat. *Exp. Brain Res.* 1996; **110**: 55–61.
49. Jankowska E, Roberts WJ. An electrophysiological demonstration of the axonal projections of single spinal interneurons in the cat. *J. Physiol.* 1972; **222**: 597–622.
50. Verberne AJ, Lewis SJ, Worland PJ *et al.* Medial prefrontal cortical lesions modulate baroreflex sensitivity in the rat. *Brain Res.* 1987; **426**: 243–9.
51. Verberne AJ, Lewis SJ, Jarrott B, Louis WJ. Medial prefrontal cortical lesions and baroreceptor heart rate reflex sensitivity in the spontaneously hypertensive rat. *J. Hypertens.* 1988; **6**: 123–7.
52. Guyenet PG, Filtz TM, Donaldson SR. Role of excitatory amino acids in rat vagal and sympathetic baroreflexes. *Brain Res.* 1987; **407**: 272–84.
53. Torrealba F, Muller C. Glutamate immunoreactivity of insular cortex afferents to the nucleus tractus solitarius in the rat: A quantitative electron microscopic study. *Neuroscience* 1996; **71**: 77–87.
54. Way M, Cechetto DF. Neurotransmitters in the lateral hypothalamic area (LHA) mediating cardiovascular responses from the infralimbic cortex (ILC). *Soc. Neurosci.* 1996; **22**: 391P.
55. Hardy SGP. Anatomical data supporting the concept of prefrontal influences upon hypothalamo-medullary relays in the rat. *Neurosci. Lett.* 1994; **169**: 17–20.
56. Hardy SGP, Mack SM. Brainstem mediation of prefrontal stimulus-produced hypotension. *Exp. Brain Res.* 1990; **79**: 393–9.
57. Verberne AJM. Medullary sympathoexcitatory neurons are inhibited by activation of the medial prefrontal cortex in the rat. *Am. J. Physiol.* 1996; **270**: R713–9.
58. Dampney RAL. The subretrofacial vasomotor nucleus: Anatomical, chemical and pharmacological properties and role in cardiovascular regulation. *Prog. Neurobiol.* 1994; **42**: 197–227.
59. van Bockstaele EJ, Pieribone VA, Aston-Jones G. Diverse afferents converge on the nucleus paragigantocellularis in the rat ventrolateral medulla: retrograde and anterograde tracing studies. *J. Comp. Neurol.* 1989; **290**: 561–84.
60. Korte SM, Jaarsma D, Luiten PGM, Bohus B. Mesencephalic cuneiform nucleus and its ascending and descending projections serve stress-related cardiovascular responses in the rat. *J. Auton. Nerv. Syst.* 1992; **41**: 157–76.
61. Edwards SB. Autoradiographic studies of the projections of the mid-brain reticular formation: Descending projections of nucleus cuneiformis. *J. Comp. Neurol.* 1975; **161**: 341–58.
62. Richter RC, Behbehani MM. Evidence for glutamic acid as a possible neurotransmitter between the mesencephalic nucleus cuneiformis and the medullary nucleus raphe magnus in the lightly anesthetized rat. *Brain Res.* 1991; **544**: 279–86.
63. Beart PM, Summers RJ, Stephenson JA, Cook CJ, Christie MJ. Excitatory amino acid projections to the periaqueductal gray in the rat: A retrograde transport study utilizing D[3H]aspartate and [3H]GABA. *Neuroscience* 1990; **34**: 163–76.
64. Beitz AJ. Possible origin of glutamatergic projections to the midbrain periaqueductal gray and deep layer of the superior colliculus of the rat. *Brain Res. Bull.* 1989; **23**: 25–35.
65. Bernard JF, Puschanski M, Besson JM. Afferents and efferents of the rat cuneiform nucleus: An anatomical study with reference to pain transmission. *Brain Res.* 1989; **490**: 181–5.
66. Redgrave P, Dean P, Mitchell IJ, Odekunle A, Clark A. The projection from superior colliculus to cuneiform area in the rat. I. Anatomical studies. *Exp. Brain Res.* 1988; **72**: 611–25.
67. Keay KA, Dean P, Redgrave P. N-Methyl-D-aspartate (NMDA) evoked changes in blood pressure and heart rate from the rat superior colliculus. *Exp. Brain Res.* 1990; **80**: 148–56.
68. Versteeg CAM, Bohus B, De Jong W. Attenuation by arginine and desglycinamide-lysine-vasopressin of a centrally evoked pressor response. *J. Auton. Nerv. Syst.* 1982; **6**: 253–62.
69. Versteeg CAM, Bohus B, De Jong W. Inhibition of centrally evoked pressor responses by neurohypophysial peptides and their fragments. *Neuropharmacology* 1982; **21**: 1359–64.
70. Morgan JJ, Curran T. Stimulus-transcription coupling in neurons: Role of cellular immediate-early genes. *Trends Neurosci.* 1989; **12**: 459–62.
71. Verberne AJM. Cuneiform nucleus stimulation produces activation of medullary sympathoexcitatory neurons in rats. *Am. J. Physiol.* 1995; **268**: R752–8.
72. Morrison SF, Milner TA, Reis DJ. Reticulospinal vasomotor neurons of

- the rat rostral ventrolateral medulla: relationship to sympathetic nerve activity and the C1 adrenergic cell group. *J. Neurosci.* 1988; **8**: 1286–301.
73. Huangfu DH, Hwang LJ, Riley TA, Guyenet PG. Role of serotonin and catecholamines in sympathetic responses evoked by stimulation of rostral medulla. *Am. J. Physiol.* 1994; **266**: R338–52.
74. Lam W, Verberne AJM. Cuneiform nucleus stimulation-induced sympathoexcitation: Role of adrenoceptors, excitatory amino acid and serotonin receptors in rat spinal cord. *Brain Res.* 1997 (in press).
75. Morrison SF. Raphe pallidus excites a unique class of sympathetic preganglionic neurons. *Am. J. Physiol.* 1993; **265**: R82–9.
76. Lam W, Gundlach AL, Verberne AJM. Neuronal activation in the fore-brain following electrical stimulation of the cuneiform nucleus in the rat: Hypothalamic expression of c-fos and NGFI-A messenger RNA. *Neuroscience* 1997; **78**: 1069–85.
77. Lam W, Gundlach AL, Verberne AJM. Increased nerve growth factor inducible-A gene and c-fos messenger RNA levels in the rat midbrain and hindbrain associated with the cardiovascular response to electrical stimulation of the mesencephalic cuneiform nucleus. *Neuroscience* 1996; **71**: 193–211.
78. Zbrozyna AW, Westwood DM. Inhibitory control of the defence–aggression reaction including its cardiovascular components. *Acta Neurobiol. Exp.* 1993; **53**: 209–13.
79. Zemlan FP, Bohbehani MM. Nucleus cuneiformis and pain modulation: Anatomy and behavioral pharmacology. *Brain Res.* 1988; **453**: 89–102.
80. Iwamoto GA, Wappel SM, Fox GM, Buetow KA, Waldrop TG. Identification of diencephalic and brainstem cardiorespiratory areas activated during exercise. *Brain Res.* 1996; **726**: 109–22.